

Review

Spirulina phenolic compounds: natural food additives with antimicrobial properties

¹*Metekia, W. A., ¹Ulusoy, B. H. and ²Habte-Tsion, H.-M.

¹Department of Food Hygiene and Technology, Faculty of Veterinary Medicine, Near East University, Near East Boulevard, 99138 Nicosia, Turkish Republic of Northern Cyprus

²University of Maine - Cooperative Extension and Aquaculture Research Institute, 5751 Murray Hall, 23 Flagstaff Road, Orono, ME 04469, United States

Article history

Received: 7 July 2020

Received in revised form:

8 April 2021

Accepted:

10 May 2021

Keywords

food safety,
fish,
seafood,
Spirulina,
phenolic,
antimicrobial,
additive

Abstract

Food safety is a scientific discipline to ensure consumers' safety and prevent food-related harms, hazards, or risks along the entire food supply chain. Although fish and seafood products are the best means for securing food and nutrition in a population, they are also highly perishable, being vulnerable to pathogenic bacteria and fungi. This review thus aimed to provide updated scientific information on the role of the phenolic compounds of *Spirulina* as food additives, and their antimicrobial activities in fish and seafood products, in a food safety context. Recent applications of *Spirulina* phenolic compounds showed good results in contaminated fish and seafood products. Different studies have concluded that *Spirulina* phenolic compounds such as polyphenols, C-phycoerythrin, γ -linolenic acid, fatty acids, and the combination of lauric and palmitoleic acids have antimicrobial activity in eliminating and/or controlling the growth of pathogenic bacteria such as *Escherichia coli*, *Pseudomonas aeruginosa*, and *Bacillus subtilis*, and fungi such as *Aspergillus flavus* and *Aspergillus niger*. Overall, *Spirulina* can be considered as an emerging natural food additive with antimicrobial activities against pathogenic fungi and bacteria.

Abbreviations:

A. flavus, *Aspergillus flavus*; *A. niger*, *Aspergillus niger*; *A. parasiticus*, *Aspergillus parasiticus*; ANSES, French Agency for Food, Environmental and Occupational Health and Safety; *A. sessilis*, *Alternanthera sessilis*; *B. subtilis*, *Bacillus subtilis*; *C. cibarius*, *Cantharellus cibarius*; *C. tora*, *Cassia tora*; *C. versicolor*, *Coriolus versicolor*; *E. coli*, *Escherichia coli*; FAO, Food and Agriculture Organization; GAC, gallic acid equivalents; *L. deliciosus*, *Lactarius deliciosus*; *L. pyrogalus*, *Lactarius pyrogalus*; *M. luteus*, *Micrococcus luteus*; *P. aeruginosa*, *Pseudomonas aeruginosa*; *P. oleracea*, *Portulaca oleracea*; QE, quercetin equivalents; *S. aureus*, *Staphylococcus aureus*; *S. platensis*, *Spirulina platensis*; *A. platensis*, *Arthrospira platensis*; *S. maxima*, *Spirulina maxima*; TPC, total phenolic content; TFC, total flavonoid content; *V. vulnificus*, *Vibrio vulnificus*; WHO, World Health Organization.

© All Rights Reserved

Introduction

Food safety is a scientific discipline to ensure consumers' safety and prevent food-related harms, hazards, or risks along the entire food supply chain, and it should be observed "from farm to fork" (WHO, 1997; FDA, 2020). Fish and seafood products are the main sources of healthy nutrition all over the world; more than 4.5 billion of the world's population get at least 15% of their animal protein intake from fish (Béné *et al.*, 2015; FAO, 2016). However, fish and seafood products are the most perishable foodstuffs. Therefore, monitoring, inspection, and quality control are the key agendas in the fishery industry.

Microbial contamination of fish and seafood products occurs after catching. During processing and postharvest, biochemical changes cause spoilage in these food products very rapidly (Møretro *et al.*, 2016; Jia *et al.*, 2019; FDA, 2020). In aquaculture, fungal contamination such as *A. flavus* has been reported in fish feed and the fish itself (Mohamed *et al.*, 2017). Another study on smoked fish reported that fungal contamination is due to unhygienic handling, filleting, and storage (Akwuobu *et al.*, 2019). Fish contamination may arise from the culturing environment such as the water used (Hennersdorf *et al.*, 2016). For example, fungi may contaminate the fish through water, soil sediments, or plant remains.

*Corresponding author.
Email: wublivelygib@gmail.com

Aspergillus and *Penicillium* have been reported as the most common fungi related to sediments and biofilms in water bodies (Gonçalves *et al.*, 2006). Foyosal and Lisa (2018) reported different *Bacillus* spp. from soil sediment, whereas other studies reported *Vibrio* spp., *Photobacterium* spp., and *Flavobacterium* spp. from different farmed fish species (Hennersdorf *et al.*, 2016). *Vibrio* spp., *Listeria monocytogenes*, *Yersinia* spp., *Salmonella* spp., and *Clostridium botulinum* have also been reported from fish and seafood products (Terentjeva *et al.*, 2015; Novoslavskij *et al.*, 2016; Don *et al.*, 2020).

Fish contaminated with fungi that could produce mycotoxins are harmful to human health (Mohamed *et al.*, 2017; Akwuobu *et al.*, 2019). Due to this, in many countries, fish and seafood products are traditionally processed and preserved to allow for long-term storage without spoilage. Preservation is mainly performed by reducing the water content in the fish, along with high and low temperature treatments, as well as combined measures (Nithin *et al.*, 2020; Tsironi *et al.*, 2020) to control and eliminate toxic bacteria such as *E. coli*, *P. aeruginosa*, and *B. subtilis*, and the growth of moulds and yeasts such as *A. niger* (El-Sheekh *et al.*, 2014).

Many studies have shown that extracted *Spirulina* phenolic compounds exhibit good results in controlling certain toxic bacteria and fungi, and could serve as an antimicrobial agent in fish and seafood products (El-Sheekh *et al.*, 2014; Pugazhendhi *et al.*, 2015; Usharani *et al.*, 2015; Pagnussatt *et al.*, 2016). Therefore, this review explores the up-to-date scientific knowledge and future perspectives regarding the antimicrobial role of *Spirulina* phenolic compounds in fish and seafood products, in a food safety context.

What is Spirulina (Arthrospira)?

Spirulina is a Gram-negative free-floating filamentous cyanobacterium capable of photosynthesis, having robust cell wall, of blue green coloration (hence the name *cyano*), and non-toxic (Fedor, 2011; Koru, 2012; Usharani *et al.*, 2012; ANSES, 2017). Prior to 1962, they were considered a eukaryotic alga (blue green algae) until they were reclassified as a prokaryote (domain Bacteria). Taxonomically, two genera are currently accepted; *Spirulina* (various species) and *Arthrospira* (important species include *A. platensis* and *A. maxima*) (Belay, 2008; ANSES, 2017).

Spirulina can be sourced and harvested from two sources; the first one is natural warm alkaline water or lakes found in African countries (Ethiopia,

Chad, and Tunisia), Latin American countries (Mexico and Peru), and Southern Asian countries (India, Sri Lanka, and Thailand) (Belay, 2008; ANSES, 2017; Assaye *et al.*, 2018; Karssa *et al.*, 2018; IOC, 2011), and the second one is artificial aquaculture ponds (Moorhead *et al.*, 2011; ANSES, 2017).

Across the world, *Spirulina* is primarily known for its high nutritional value, and considered as a miracle / super food (Moorhead *et al.*, 2011; Tidjani *et al.*, 2018; Uddin *et al.*, 2018; Jung *et al.*, 2019), functional food and dietary supplement (ANSES, 2017; Panjiar *et al.*, 2017), feed supplement (Yusuf *et al.*, 2016), technofunctional ingredient in meat products to improve meat quality (de Medeiros *et al.*, 2020), and protein and vitamin supplement for fish farming industries (Usharani *et al.*, 2012; Ragaza *et al.*, 2020). *Spirulina* is rich in micro- and macronutrients (Moorhead *et al.*, 2011; ANSES, 2017), and used as an ingredient in the production of valuable innovative foods in the food industries (Lafarga *et al.*, 2020). It is also considered as a future food for mankind around the globe (Belay, 2008; Soni *et al.*, 2017; Mathur, 2019). Throughout history, the Aztecs in Mexico and Chad communities in tropical Africa have eaten the naturally harvested *Spirulina* in a dried cake form (Ahsan *et al.*, 2008; Tidjani *et al.*, 2018; Lafarga *et al.*, 2020).

Spirulina contains desirable chemical compositions such as proteins, carbohydrates (e.g., sulphated polysaccharides), essential amino acids, minerals, essential fatty acids (e.g., γ -linoleic acid), vitamins, and pigments (e.g., C-phycocyanin) (Belay, 2008; Thomas, 2010; IOC, 2011; Lafarga *et al.*, 2020).

Phenols and polyphenols of Spirulina

In organic chemistry, phenols or polyphenols are defined as compounds that have one or more hydroxy substituents in an aromatic ring, such as structural compounds (e.g., esters, methyl ethers, glycosides) (Lattanzio, 2013). Together, these phenolic compounds range from simple phenols to polyphenols and can be derived from plants and microorganisms including *Spirulina* (Al-Dhabi and Valan Arasu, 2016; Hussain *et al.*, 2019; Olszewska *et al.*, 2020). Most phenolics have two or more classes of hydroxyl, and are biologically active, having antimicrobial and/or antioxidant properties. Phenolics occur widely in plant sources, and are consumed daily by the global population, as illustrated in Table 1. Polyphenols have different categories based on their food sources, which include flavonoids, phenolic acids, lignans, and stilbenes.

Table 1. Phenolic and flavonoid contents from selected foods.

Author	Article title	Studied food	Total phenolic content (mg GAE/g)	Total flavonoid content (μg QE/g)
El-Baky <i>et al.</i> (2009)	Production of phenolic compounds from <i>Spirulina maxima</i> microalgae and its protective effects	<i>Spirulina maxima</i>	12.94 ± 0.93	4.65 ± 0.14
Gurnani <i>et al.</i> (2016)	Chemical composition, total phenolic and flavonoid contents, and <i>in vitro</i> antimicrobial and antioxidant activities of crude extracts from red chilli seeds (<i>Capsicum frutescens</i> L.)	Chili seed	7.95 - 26.15	4.64 - 12.84
Jang <i>et al.</i> (2018)	Antioxidant and antimicrobial activities of fresh garlic and aged garlic by-products extracted with different solvents	Garlic	47.58 ± 5.27	338.04 ± 1.60^a
Alexandre <i>et al.</i> (2019)	Antimicrobial activity of pomegranate peel extracts performed by high pressure and enzymatic assisted extraction	Pomegranate	219 ± 2.6^b and 211 ± 0.7	No report
Ayvaz <i>et al.</i> (2019)	Phenolic profile of three wild edible mushroom extracts from Ordu, Turkey and their antioxidant properties, enzyme inhibitory activities	Mushroom	2.27 to 5.34 ± 0.55	5.02 to 17.54 ± 1.45
Papalia <i>et al.</i> (2019)	Impact of different storage methods on bioactive compounds in <i>Arthrospira platensis</i> biomass	<i>Arthrospira platensis</i>	15.77 ± 1.10^b	20.82 ± 0.08^c
Seghiri <i>et al.</i> (2019)	Functional composition, nutritional properties, and biological activities of Moroccan <i>Spirulina</i> microalga	<i>Spirulina</i>	4.19 ± 0.21	15.60 ± 2.74

^aaged garlic powder extracted with distilled water; ^bhigh pressure extraction condition; and ^ctotal phenol and flavonoid of fresh *A. platensis*.

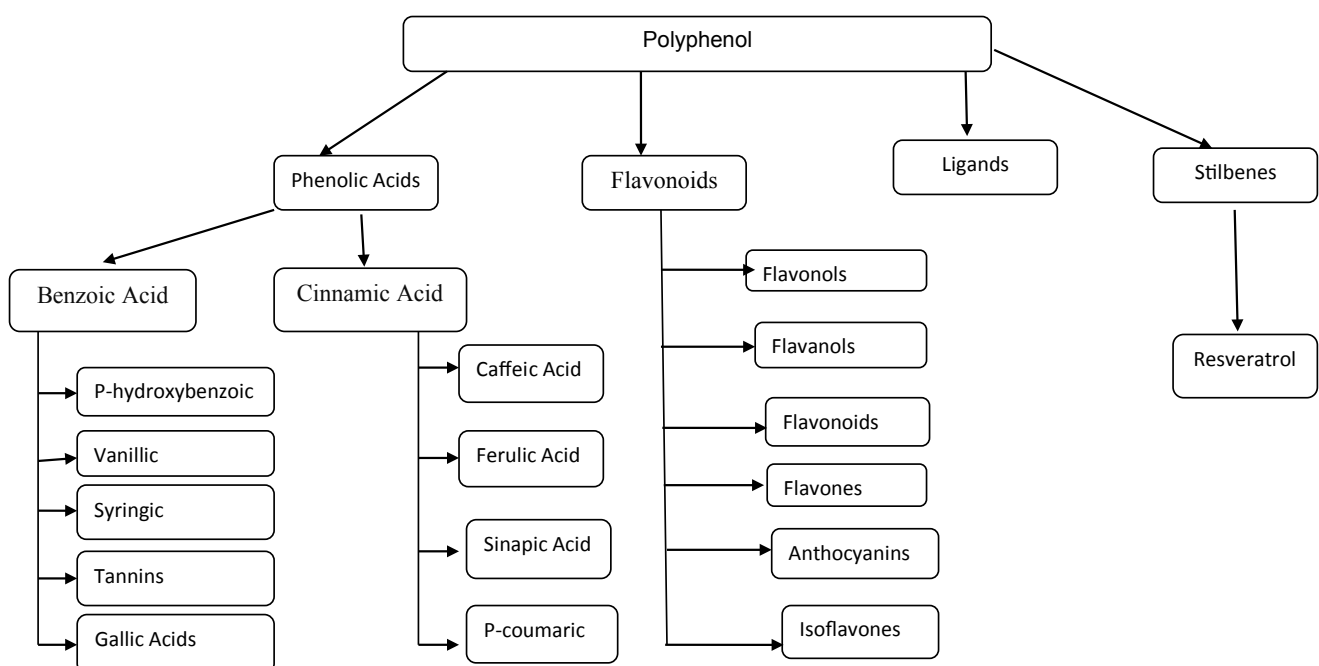


Figure 1. Categories of polyphenols.

Figure 1 illustrates the categories of polyphenols (Becerra-Herrera *et al.*, 2014; de Medina *et al.*, 2015; Papalia *et al.*, 2019).

The functions of *Spirulina* polyphenols are not only limited to antimicrobial and antimutagenic activities, but also anticancer, antiaging, anti-inflammation, lung defensive; neuroprotective, cardioprotective, hepatoprotective, and many more as illustrated in Figure 2. Furthermore, different studies have indicated their antimicrobial roles against different foodborne pathogens (Ghomari *et al.*, 2019; Papalia *et al.*, 2019; Polverari *et al.*, 2019), antioxidative roles (Salamatullah, 2014; Irakli *et al.*, 2018; Papalia *et al.*, 2019; Kukharensko *et al.*, 2020), antifungal and antimycotoxin roles (Shishido *et al.*, 2015; Christ-Ribeiro *et al.*, 2019), cardiovascular health promoting properties (Huang *et al.*, 2018), control and elimination of life-threatening viruses (Carrera-González *et al.*, 2013; Martínez-Martos *et al.*, 2014; Mathur, 2019), neural protection and boosting the immune system (Martínez-Martos *et al.*, 2014; Kumar *et al.*, 2017), antidiabetes, antihypertensive, and anti-obesity roles (Hu *et al.*, 2019; Lafarga *et al.*, 2020), anticancer properties (Jung *et al.*, 2019; Marková *et al.*, 2020), and the prevention of retinal neurodegeneration and improvement of eyesight (Okamoto *et al.*, 2019). Another study showed the chemopreventive role of *Spirulina* water extract solution which revealed good results in a human lung cancer A549 cell line with a considerable reduction of cancer cell viability and multiplication (Czerwonka *et al.*, 2018).

The quantity and efficiency of phenols, polyphenols, C-phycoyanin, and other bioactive compounds like γ -linolenic acid and carotenoids depend on the growth medium and condition of *Spirulina* (Sassano *et al.*, 2014; Gupta *et al.*, 2018; Papalia *et al.*, 2019), light intensity (Kepekçi and Saygideger, 2012), storage (Papalia *et al.*, 2019), processing and extraction methods, and the solvents used (Salamatullah, 2014; Papalia *et al.*, 2019). According to Papalia *et al.* (2019), the highest quantity of phenols, phycoyanin-C, and ascorbic acid was found in fresh *Spirulina* biomass, and the highest total flavonoid content was found in freeze-dried *Spirulina* biomass. On the other hand, the highest antimicrobial activity was found in both fresh and freeze-dried biomass due to the high carotenoid concentration, thus indicating the high concentration of phenolic compounds present in *Spirulina*.

Antimicrobial effect of Spirulina phenolic compounds

Antimicrobials are natural or man-made substances that can be used to control, kill, and/or inhibit the development and growth of microorganisms including viruses, bacteria, fungi, and protozoa (de Medina *et al.*, 2015; Sherrard *et al.*, 2014). *Spirulina* is known globally as a dietary supplement for human consumption, but in recent times, it has also been recognised as a good antimicrobial compound due to its ability to control and inhibit the development or growth of foodborne pathogens and

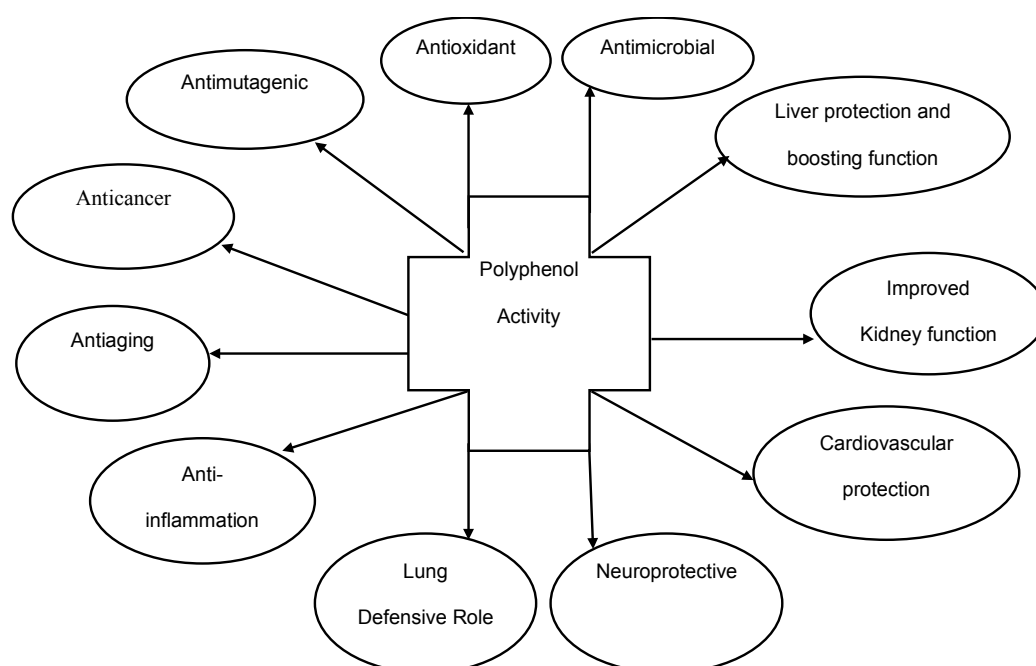


Figure 2. Polyphenol activities of *Spirulina* spp.

food spoilage microorganisms in the food industry (Duda-Chodak, 2013; Seghiri *et al.*, 2019; Elshouny *et al.*, 2020; Martelli *et al.*, 2020). Different studies reported the antimicrobial effect of *Spirulina* on the growth of different bacteria, yeasts, and moulds including *Alicyclobacillus acidoterrestris*, *Aspergillus flavus*, *Aspergillus niger*, *Bacillus subtilis*, *Cladosporium*, *Geotrichum*, *Micrococcus luteus*, *Penicillium*, and *Rhodotorula* (Souza *et al.*, 2011; Duda-Chodak, 2013).

The need to produce safe foods has encouraged the exploration of natural ingredients that possess antimicrobial activity (Souza *et al.*, 2011; Jung *et al.*, 2019; Martelli *et al.*, 2020). Some of these natural ingredients can be obtained from *Spirulina*. *Spirulina* phenolic compound extracts have shown excellent antimicrobial activities (Sun *et al.*, 2016; Swain *et al.*, 2017; Jung *et al.*, 2019). *Spirulina platensis* methanolic extract (1.15 mg phenolic compound per gram of *S. platensis* biomass) has shown antifungal effect against *A. flavus* at 54 µg/mL minimum inhibitory concentration (MIC), four times lower than the control (Souza *et al.*, 2011). Parisi *et al.* (2009) reported that *S. platensis* phenolic compound extract inhibited *S. aureus* at 47.5 mg/mL MIC. Scaglioni *et al.* (2019) reported 50% inhibition using 33.9 µg/mL *S. platensis* phenolic compound extract against *Fusarium*. Other studies reported good inhibition results of *S. platensis* phenolic compound extract against *E. coli*, *P. aeruginosa*, *B. subtilis*, *V. vulnificus*, *A. flavus*, and *A. niger* (El-Sheekh *et al.*, 2014; Rajasekar *et al.*, 2019; Seghiri *et al.*, 2019). C-phycoerythrin from *S. platensis* inhibited drug-resistant bacteria such as *E. coli*, *K. pneumoniae*, *P. aeruginosa*, and *S. aureus* (Sarada *et al.*, 2011).

Another study regarding the inhibition success of *Spirulina* phenolic compound extract against fungus was carried out, and the application of 8% liposomal phenolic compound was found more effective, with 90% inhibition, than the free extract solution, with 74% inhibition (Pugazhendhi *et al.*, 2015). Seghiri *et al.* (2019) reported that the methanolic extract of *Spirulina* phenolic compound exhibited a good result against pathogenic bacteria and fungi. It was assumed that the cell concentrate of *S. platensis* has the suitable ability to control the development and growth of pathogenic bacteria and fungi including *B. subtilis*, *S. aureus*, *E. coli*, *M. luteus*, *A. flavus*, *A. niger*, and *C. versicolor* (Usharani *et al.*, 2015; Pugazhendhi *et al.*, 2015; Seghiri *et al.*, 2019).

Other researchers reported that the extract of *S. platensis* is active for the control and inhibition of

fungi and bacteria, and serves as a good source of antimicrobial activity due to the bioactive compounds like γ -linolenic acid, fatty acids, and interaction of lauric and palmitoleic acids (Mendiola *et al.*, 2007; Ramadan *et al.*, 2008; El-Sheekh *et al.*, 2014; Yang *et al.*, 2019). Other studies reported that the antibacterial activity of C-phycoerythrin from *S. platensis* was amplified at higher concentrations, specifically 400 µL/disk and above (Safari *et al.*, 2019). This is possibly due to the resistance of those bacterial strains to the extract of C-phycoerythrin differing from one species to another. As concluded in a study conducted by Safari *et al.* (2019), *Listeria* and *Streptococcus* were the most susceptible and resistant bacterial isolates to C-phycoerythrin, respectively. The researchers' finding also highlighted that C-phycoerythrin from *S. platensis* has excellent antimicrobial activity, and it can be used as a natural antimicrobial in different food commodities, mainly foods with high lipids, as well as in cosmetic products. Both antioxidant and antibacterial activities of C-phycoerythrin decrease in long-term storage at -18°C, most likely due to the effect of the freezing temperature on the protein structure of C-phycoerythrin (Safari *et al.*, 2019; Usharani *et al.*, 2015).

Conclusion

Fish and seafood products are the best means of food and nutrition security. At the same time, however, they are highly perishable, being vulnerable to pathogenic microorganisms including *E. coli*, *P. aeruginosa*, *B. subtilis*, *A. flavus*, and *A. niger*. Currently, *Spirulina* is known as a dietary supplement, and its bioactive compound extracts are known for their antimicrobial activity towards pathogenic bacteria and fungi. Promoting this emerging natural compound in fish and fish products will save both our food and the public from food-related risks. *Spirulina* bioactive compounds include polyphenols, phycoerythrin, and fatty acids. Flavonoids and phenolic acids from the polyphenol category could be promising. Future perspectives of *Spirulina* should include issues such as multifunctional study of polyphenols including antimicrobial agent development, drug development, and application in animal and fish feed production. Implications for climate mitigation measures should also be further elucidated and studied.

Acknowledgement

Special thanks to Dr. Amha Belay, a senior algal researcher and resource person on *Spirulina*, for the technical assistance rendered.

References

- Ahsan, M., Habib, B., Parvin, M., Huntington, T. and Hasan, M. 2008. A review on culture, production and use of *Spirulina* as food for humans and feeds for domestic animals. Rome: Food and Agriculture Organization (FAO).
- Akwuobu, C. A., Antiev, W. S. and Ofukwu, R. A. P. 2019. Fungal contaminants of smoke-dried fish sold in open markets in Makurdi, Benue state, north-central Nigeria. *Food and Nutrition Sciences* 10(3): 290-297.
- Al-Dhabi, N. A. and Valan Arasu, M. 2016. Quantification of phytochemicals from commercial *Spirulina* products and their antioxidant activities. *Evidence-Based Complementary and Alternative Medicine* 2016: article ID 7631864.
- Alexandre, E. M., Silva, S., Santos, S. A., Silvestre, A. J., Duarte, M. F., Saraiva, J. A. and Pintado, M. 2019. Antimicrobial activity of pomegranate peel extracts performed by high pressure and enzymatic assisted extraction. *Food Research International* 115: 167-176.
- Assaye, H., Belay, A., Desse, G. and Gray, D. 2018. Seasonal variation in the nutrient profile of *Arthrospira fusiformis* biomass harvested from an Ethiopian soda lake, Lake Chitu. *Journal of Applied Phycology* 30(3): 1597-1606.
- Ayvaz, M. Ç., Aksu, F. and Kır, F. 2019. Phenolic profile of three wild edible mushroom extracts from Ordu, Turkey and their antioxidant properties, enzyme inhibitory activities. *British Food Journal* 121(6): 1248-1260.
- Becerra-Herrera, M., Sánchez-Astudillo, M., Beltrán, R. and Sayago, A. 2014. Determination of phenolic compounds in olive oil: new method based on liquid-liquid micro extraction and ultra-high performance liquid chromatography-triple-quadrupole mass spectrometry. *LWT - Food Science and Technology* 57(1): 49-57.
- Belay, A. 2008. *Spirulina (Arthrospira)*: production and quality assurance. In Gershwin, M. E. and Belay, A. (eds). *Spirulina in Human Nutrition and Health* (1st ed). United States: CRC Press.
- Béné, C., Barange, M., Subasinghe, R., Pinstrup-Andersen, P., Merino, G., Hemre, G.-I. and Williams, M. 2015. Feeding 9 billion by 2050 - putting fish back on the menu. *Food Security* 7(2): 261-274.
- Carrera-González, M., Ramírez-Expósito, M., Mayas, M. and Martínez-Martos, J. 2013. Protective role of oleuropein and its metabolite hydroxytyrosol on cancer. *Trends in Food Science and Technology* 31(2): 92-99.
- Christ-Ribeiro, A., Graça, C., Kupski, L., Badiale-Furlong, E. and De Souza-Soares, L. A. 2019. Cytotoxicity, antifungal and anti-mycotoxins effects of phenolic compounds from fermented rice bran and *Spirulina* sp. *Process Biochemistry* 80: 190-196.
- Czerwonka, A., Kaławaj, K., Sławińska-Brych, A., Lemieszek, M. K., Bartnik, M., Wojtanowski, K. K., ... and Rzeski, W. 2018. Anticancer effect of the water extract of a commercial *Spirulina (Arthrospira platensis)* product on the human lung cancer A549 cell line. *Biomedicine and Pharmacotherapy* 106: 292-302.
- de Medeiros, V. P. B., Pimentel, T. C., Sant'Ana, A. S. and Magnani, M., 2020. Microalgae in the meat processing chain: feed for animal production or source of techno-functional ingredients. *Current Opinion in Food Science* 37: 125-134.
- de Medina, V. S., Priego-Capote, F. and De Castro, M. D. L. 2015. Characterization of monovarietal virgin olive oils by phenols profiling. *Talanta* 132: 424-432.
- Don, S., Ammini, P., Nayak, B. B. and Kumar, S. H. 2020. Survival behaviour of *Salmonella enterica* in fish and shrimp at different conditions of storage. *LWT - Food Science and Technology* 132: article ID 109795.
- Duda-Chodak, A. 2013. Impact of water extracts of *Spirulina* (WES) on bacteria, yeasts and molds. *Acta Scientiarum Polonorum Technologia Alimentaria* 12(1): 33-39.
- El-Baky, H., El Baz, F. K. and El-Baroty, G. S. 2009. Production of phenolic compounds from *Spirulina maxima* microalgae and its protective effects *in vitro* toward hepatotoxicity model. *African Journal of Pharmacy and Pharmacology* 3(4): 133-139.
- El-Sheekh, M. M., Daboor, S. M., Swelim, M. A. and Mohamed, S. 2014. Production and characterization of antimicrobial active substance from *Spirulina platensis*. *Iranian Journal of Microbiology* 6(2): 112-119.
- Elshouny, W. A. E. F., El-Sheekh, M. M., Sabae, S. Z., Khalil, M. A. and Badr, H. M. 2020. Antimicrobial activity of *Spirulina platensis* against aquatic bacterial isolates. *Journal of Microbiology, Biotechnology and Food Sciences* 9(6): 1203-1208.
- Fedor, K. 2011. *Arthrospira platensis*. Retrieved from website: http://bioweb.uwlax.edu/bio203/2011/fedor_kara/
- Food and Agriculture Organization (FAO). 2016.

- Methods for estimating comparable rates of food insecurity experienced by adults throughout the world. Rome: FAO.
- Food and Drug Administration (FDA). 2020. Fish and fishery products hazards and controls. 4th ed. United States: FDA.
- Foyosal, M. J. and Lisa, A. K. 2018. Isolation and characterization of *Bacillus* sp. strain BC01 from soil displaying potent antagonistic activity against plant and fish pathogenic fungi and bacteria. *Journal of Genetic Engineering and Biotechnology* 16(2): 387-392.
- French Agency for Food, Environmental and Occupational Health and Safety (ANSES). 2017. Opinion of the French Agency for Food, Environmental and Occupational Health and Safety on the risks associated with the consumption of food supplements containing *Spirulina*. Retrieved from ANSES website: <https://www.anses.fr/en/system/files/NUT2014-SA0096EN.pdf>
- Ghomari, O., Sounni, F., Massaoudi, Y., Ghanam, J., Kaitouni, L. B. D., Merzouki, M. and Benlemlih, M. 2019. Phenolic profile (HPLC-UV) of olive leaves according to extraction procedure and assessment of antibacterial activity. *Biotechnology Reports* 23: article ID e00347.
- Gonçalves, A. B., Paterson, R. R. M. and Lima, N. 2006. Survey and significance of filamentous fungi from tap water. *International Journal of Hygiene and Environmental Health* 209(3): 257-264.
- Gupta, A., Mohan, D., Saxena, R. K. and Singh, S. 2018. Phototrophic cultivation of NaCl-tolerant mutant of *Spirulina platensis* for enhanced C-phycoerythrin production under optimized culture conditions and its dynamic modeling. *Journal of Phycology* 54(1): 44-55.
- Gurnani, N., Gupta, M., Mehta, D. and Mehta, B. K. 2016. Chemical composition, total phenolic and flavonoid contents, and *in vitro* antimicrobial and antioxidant activities of crude extracts from red chilli seeds (*Capsicum frutescens* L.). *Journal of Taibah University for Science* 10(4): 462-470.
- Hennersdorf, P., Kleinertz, S., Theisen, S., Abdul-Aziz, M. A., Mrotzek, G., Palm, H. W. and Saluz, H. P. 2016. Microbial diversity and parasitic load in tropical fish of different environmental conditions. *PLoS One* 11(3): article ID e0151594.
- Hu, S., Fan, X., Qi, P. and Zhang, X. 2019. Identification of anti-diabetes peptides from *Spirulina platensis*. *Journal of Functional Foods* 56: 333-341.
- Huang, H., Liao, D., Pu, R. and Cui, Y. 2018. Quantifying the effects of *Spirulina* supplementation on plasma lipid and glucose concentrations, body weight, and blood pressure. *Diabetes, Metabolic Syndrome and Obesity - Targets and Therapy* 11: 729-742.
- Hussain, M. B., Hassan, S., Waheed, M., Javed, A., Farooq, M. A. and Tahir, A. 2019. Bioavailability and metabolic pathway of phenolic compounds. In Soto-Hernández, M., García-Mateos, R. and Palma-Tenango, M. (eds). *Plant Physiological Aspects of Phenolic Compounds*. United Kingdom: IntechOpen.
- Indian Ocean Commission (IOC). 2011. *Spirulina - a livelihood and a business venture*. Mauritius: IOC.
- Irakli, M., Chatzopoulou, P. and Ekateriniadou, L. 2018. Optimization of ultrasound-assisted extraction of phenolic compounds: oleuropein, phenolic acids, phenolic alcohols and flavonoids from olive leaves and evaluation of its antioxidant activities. *Industrial Crops and Products* 124: 382-388.
- Jang, H.-J., Lee, H.-J., Yoon, D.-K., Ji, D.-S., Kim, J.-H. and Lee, C.-H. 2018. Antioxidant and antimicrobial activities of fresh garlic and aged garlic by-products extracted with different solvents. *Food Science and Biotechnology* 27(1): 219-225.
- Jia, S., Li, Y., Zhuang, S., Sun, X., Zhang, L., Shi, J., ... and Luo, Y., 2019. Biochemical changes induced by dominant bacteria in chill-stored silver carp (*Hypophthalmichthys molitrix*) and GC-IMS identification of volatile organic compounds. *Food Microbiology* 84: article ID 103248.
- Jung, F., Krüger-Genge, A., Waldeck, P. and Küpper, J. H. 2019. *Spirulina platensis*, a super food? *Journal of Cellular Biotechnology* 5(1): 43-54.
- Karssa, T., Papini, A. and Kasan, N. A. 2018. Cultivation of *Arthrospira* strains in tropical conditions, with particular reference to Ethiopia. *International Journal of Food Science and Nutrition Engineering* 8: 107-118.
- Kepekçi, R. A. and Saygideger, S. D. 2012. Enhancement of phenolic compound production in *Spirulina platensis* by two-step batch mode cultivation. *Journal of Applied Phycology* 24(4): 897-905.
- Koru, E. 2012. Earth food *Spirulina* (*Arthrospira*): production and quality standards. In El-Samragy, Y. (ed). *Food Additive*, p. 191-202. United Kingdom: IntechOpen.

- Kukhareenko, A., Brito, A., Yashin, Y. I., Yashin, A. Y., Kuznetsov, R. M., Markin, P. A., ... and Appolonova, S. A. 2020. Total antioxidant capacity of edible plants commonly found in East Asia and the Middle East determined by an amperometric method. *Journal of Food Measurement and Characterization* 14(2): 809-817.
- Kumar, A., Christian, P. K., Panchal, K., Guruprasad, B. and Tiwari, A. K. 2017. Supplementation of *Spirulina* (*Arthrospira platensis*) improves lifespan and locomotor activity in paraquat-sensitive *DJ-1^{Δ93}* flies, a Parkinson's disease model in *Drosophila melanogaster*. *Journal of Dietary Supplements* 14(5): 573-588.
- Lafarga, T., Fernández-Sevilla, J. M., González-López, C. and Ación-Fernández, F. G. 2020. *Spirulina* for the food and functional food industries. *Food Research International* 137: article ID 109356.
- Lattanzio, V. 2013. Phenolic compounds - introduction. In Ramawat, K. G. and Mérillon, J.-M. (eds). *Natural Products*, p. 1543-1580. Berlin: Springer.
- Marková, I., Koničková, R., Vaňková, K., Leníček, M., Kolář, M., Strnad, H., ... and Klímová, Z. 2020. Anti-angiogenic effects of the blue-green alga *Arthrospira platensis* on pancreatic cancer. *Journal of Cellular and Molecular Medicine* 24(4): 2402-2415.
- Martelli, F., Cirlini, M., Lazzi, C., Neviani, E. and Bernini, V. 2020. Edible seaweeds and *Spirulina* extract for food application: *in vitro* and *in situ* evaluation of antimicrobial activity towards foodborne pathogenic bacteria. *Foods* 9(10): article no. 1442.
- Martínez-Martos, J. M., Mayas, M. D., Carrera, P., De Saavedra, J. M. A., Sánchez-Agosta, R., Arrazola, M. and Ramírez-Expósito, M. J. 2014. Phenolic compounds oleuropein and hydroxytyrosol exert differential effects on glioma development via antioxidant defense systems. *Journal of Functional Foods* 11: 221-234.
- Mathur, M. 2019. Bioactive molecules of *Spirulina*: a food supplement. In Mérillon, J.-M. and Ramawat, K. G. (eds). *Bioactive Molecules in Food*, p. 1-22. United States: Springer.
- Mendiola, J. A., Jaime, L., Santoyo, S., Reglero, G., Cifuentes, A., Ibañez, E. and Señoráns, F. 2007. Screening of functional compounds in supercritical fluid extracts from *Spirulina platensis*. *Food Chemistry* 102(4): 1357-1367.
- Mohamed, H. M., Emeish, W. F., Braeuning, A. and Hammad, S. 2017. Detection of aflatoxin-producing fungi isolated from Nile tilapia and fish feed. *EXCLI Journal* 16: 1308-1318.
- Moorhead, K., Capelli, B. and Cysewski, G. R. 2011. *Spirulina* - nature's superfood. Hawaii: Cyano-tech Corporation.
- Møretro, T., Moen, B., Heir, E., Hansen, A. Å. and Langsrud, S. 2016. Contamination of salmon fillets and processing plants with spoilage bacteria. *International Journal of Food Microbiology* 237: 98-108.
- Nithin, C. T., Joshy, C. G., Chatterjee, N. S., Panda, S. K., Yathavamoorthi, R., Ananthanarayanan, T. R., ... and Gopal, T. K. S. 2020. Liquid smoking - a safe and convenient alternative for traditional fish smoked products. *Food Control* 113: article ID 107186.
- Novoslavskij, A., Terentjeva, M., Eizenberga, I., Valciņa, O., Bartkevičs, V. and Bērziņš, A. 2016. Major foodborne pathogens in fish and fish products: a review. *Annals of Microbiology* 66(1): 1-15.
- Okamoto, T., Kawashima, H., Osada, H., Toda, E., Homma, K., Nagai, N., ... and Ozawa, Y. 2019. Dietary *Spirulina* supplementation protects visual function from photostress by suppressing retinal neurodegeneration in mice. *Translational Vision Science and Technology* 8(6): article no. 20.
- Olszewska, M. A., Gėdas, A. and Simões, M. 2020. Antimicrobial polyphenol-rich extracts: applications and limitations in the food industry. *Food Research International* 134: article ID 109214.
- Pagnussatt, F. A., de Lima, V. R., Dora, C. L., Costa, J. A. V., Putaux, J.-L. and Badiale-Furlong, E. 2016. Assessment of the encapsulation effect of phenolic compounds from *Spirulina* sp. *LEB-18* on their antifusarium activities. *Food Chemistry* 211: 616-623.
- Panjiar, N., Mishra, S., Yadav, A. N. and Verma, P. 2017. Functional foods from cyanobacteria: an emerging source for functional food products of pharmaceutical importance. In Gupta, V. K., Treichel, H., Shapaval, V., de Oliveira, L. A. and Tuohy, M. G. (eds). *Microbial Functional Foods and Nutraceuticals*, p. 21-37. United States: Wiley.
- Papalia, T., Sidari, R. and Panuccio, M. R. 2019. Impact of different storage methods on bioactive compounds in *Arthrospira platensis* biomass. *Molecules* 24(15): article no. 2810.
- Parisi, A., Younes, S., Reinehr, C. and Colla, L. 2009. Assessment of the antibacterial activity of microalgae *Spirulina platensis*. *Journal of Basic*

- and Applied Pharmaceutical Sciences 30(3): 297-301.
- Polverari, A., Lovato, A., Vitulo, N., Vandelle, E. G. and Pignatti, A. 2019. Inhibition of virulence-related traits in *Pseudomonas syringae* pv. *actinidiae* by Gunpowder green tea extracts. *Frontiers in Microbiology* 10: article no. 2362.
- Pugazhendhi, A., Margaret Muthu Rathinam, A. and Mary Sheela, J. 2015. Antifungal activity of cell extract of *Spirulina platensis* against aflatoxin producing *Aspergillus* species. *International Journal of Current Microbiology and Applied Sciences* 4(8): 1025-1029.
- Ragaza, J. A., Hossain, M. S., Meiler, K. A., Velasquez, S. F. and Kumar, V. 2020. A review on *Spirulina*: alternative media for cultivation and nutritive value as an aquafeed. *Reviews in Aquaculture* 12(4): 2371-2395.
- Rajasekar, P., Palanisamy, S., Anjali, R., Vinosha, M., Elakkiya, M., Marudhupandi, T., ... and Prabhu, N. M. 2019. Isolation and structural characterization of sulfated polysaccharide from *Spirulina platensis* and its bioactive potential: *in vitro* antioxidant, antibacterial activity and Zebrafish growth and reproductive performance. *International Journal of Biological Macromolecules* 141: 809-821.
- Ramadan, M. F., Asker, M. M. S. and Ibrahim, Z. K. 2008. Functional bioactive compounds and biological activities. *Czech Journal of Food Sciences* 26(3): 211-222.
- Safari, R., Raftani Amiri, Z. and Esmaeilzadeh Kenari, R. 2019. Antioxidant and antibacterial activities of C-phycoerythrin from common name *Spirulina platensis*. *Iranian Journal of Fisheries Sciences* 19(4): 1911-1927.
- Salamatullah, A. 2014. Characterization of extraction methods to recover phenolic-rich antioxidants from blue green algae (*Spirulina*) using response surface approaches. United States: University of Nebraska, MSc thesis.
- Sarada, D. V., Kumar, C. S. and Rengasamy, R. 2011. Purified C-phycoerythrin from *Spirulina platensis* (Nordstedt) Geitler: a novel and potent agent against drug resistant bacteria. *World Journal of Microbiology and Biotechnology* 27(4): 779-783.
- Sassano, C. E. N., Gioielli, L. A., Converti, A., De Oliveira Moraes, I., Sato, S. and De Carvalho, J. C. M. 2014. Urea increases fed-batch growth and γ -linolenic acid production of nutritionally valuable *Arthrospira* (*Spirulina*) *platensis* cyanobacterium. *Engineering in Life Sciences* 14(5): 530-537.
- Scaglioni, P. T., Pagnussatt, F. A., Lemos, A. C., Nicolli, C. P., Del Ponte, E. M. and Badiale-Furlong, E. 2019. *Nannochloropsis* sp. and *Spirulina* sp. as a source of antifungal compounds to mitigate contamination by *Fusarium graminearum* species complex. *Current Microbiology* 76(8): 930-938.
- Seghiri, R., Kharbach, M. and Essamri, A., 2019. Functional composition, nutritional properties, and biological activities of Moroccan *Spirulina* microalga. *Journal of Food Quality* 2019: article ID 3707219.
- Sherrard, L. J., Tunney, M. M. and Elborn, J. S. 2014. Antimicrobial resistance in the respiratory microbiota of people with cystic fibrosis. *The Lancet* 384(9944): 703-713.
- Shishido, T. K., Humisto, A., Jokela, J., Liu, L., Wahlsten, M., Tamrakar, A., ... and Sivonen, K. 2015. Antifungal compounds from cyanobacteria. *Marine Drugs* 13(4): 2124-2140.
- Soni, R. A., Sudhakar, K. and Rana, R. 2017. *Spirulina* - from growth to nutritional product: a review. *Trends in Food Science and Technology* 69: 157-171.
- Souza, M. M. D., Prietto, L., Ribeiro, A. C., Souza, T. D. D. and Badiale-Furlong, E. 2011. Assessment of the antifungal activity of *Spirulina platensis* phenolic extract against *Aspergillus flavus*. *Ciência e Agrotecnologia* 35(6): 1050-1058.
- Sun, Y., Chang, R., Li, Q. and Li, B. 2016. Isolation and characterization of an antibacterial peptide from protein hydrolysates of *Spirulina platensis*. *European Food Research and Technology* 242(5): 685-692.
- Swain, S. S., Paidesetty, S. K. and Padhy, R. N. 2017. Antibacterial, antifungal and antimycobacterial compounds from cyanobacteria. *Biomedicine and Pharmacotherapy* 90: 760-776.
- Terentjeva, M., Eizenberga, I., Valciņa, O., Novoslavskij, A., Strazdiņa, V. and Bērziņš, A. 2015. Prevalence of foodborne pathogens in freshwater fish in Latvia. *Journal of Food Protection* 78(11): 2093-2098.
- Thomas, S. S. 2010. The role of parry organic *Spirulina* in health management. India: Parry Nutraceuticals.
- Tidjani, A., Alio, H. M., Aguid, M. N. and Doutoum, A. A. 2018. Study of the microbiological quality of improved *Spirulina* marketed in Chad. *Nutrition and Food Science International Journal* 4(4): 91-95.
- Tsironi, T., Houhoula, D. and Taoukis, P. 2020. Hurdle technology for fish preservation.

- Aquaculture and Fisheries 5(2): 65-71.
- Uddin, A. J., Mahbuba, S., Rahul, S., Ifaz, M. and Ahmad, H. 2018. Super food *Spirulina* (*Spirulina platensis*): prospect and scopes in Bangladesh. International Journal of Business, Social and Scientific Research 6: 51-55.
- Usharani, G., Saranraj, P. and Kanchana, D. 2012. *Spirulina* cultivation: a review. International Journal of Pharmaceutical and Biological Archives 3(6): 1327-1341.
- Usharani, G., Srinivasan, G., Sivasakthi, S. and Saranraj, P. 2015. Antimicrobial activity of *Spirulina platensis* solvent extracts against pathogenic bacteria and fungi. Advances in Biological Research 9(5): 292-298.
- World Health Organization (WHO). 1997. Risk management and food safety. Rome: WHO.
- Yang, X., Li, Y., Li, Y., Ye, D., Yuan, L., Sun, Y., ... and Hu, Q. 2019. Solid matrix-supported supercritical CO₂ enhances extraction of γ -linolenic acid from the cyanobacterium *Arthrospira* (*Spirulina*) *platensis* and bioactivity evaluation of the molecule in Zebrafish. Marine Drugs 17(4): article no. 203.
- Yusuf, M. S., Hassan, M. A., Abdel-Daim, M. M., El Nabtiti, A. S., Ahmed, A. M., Moawed, S. A., ... and Cui, H. 2016. Value added by *Spirulina platensis* in two different diets on growth performance, gut microbiota, and meat quality of Japanese quails. Veterinary World 9(11): 1287-1293.